

Impact of sodium azide on maturity, seed and tuber yields in M₁ African yam bean [*Sphenostylis stenocarpa* (Hochst ex A. Rich) Harms] generation

Solomon Tayo Akinyosoye*; Johnson Adedayo Adetumbi and Paul Chiedozie Ukachukwu

Institute of Agricultural Research and Training, Obafemi Awolowo University, Ibadan, Nigeria

*Corresponding author email: stakinyosoye@gmail.com

Mutation breeding has been recognized as a valuable supplement tool to conventional breeding and it has contributed significantly to crop improvement. This study therefore, evaluated the effects of sodium azide (NaN₃) on maturity, tuber and seed yields related traits of African yam bean (AYB) accessions, with the aim of creating genetic variation. Six promising AYB genotypes with traits of early maturity and high yield were selected from previous evaluations across agro-ecological zones in southwest Nigeria. Seeds of AYB were pre-soaked in distilled water at room temperature (25 ± 2 °C) for 6 hours. The pre-soaked seeds were then treated with different NaN₃ concentrations (0.02%, 0.04%, 0.06%, 0.08%) for 9 hours; the control was also soaked for 9 hours in distilled water. The treated seeds were sown in the field for evaluation of M₁ generation. The results obtained revealed that genotypes, NaN₃ concentrations and their interaction, showed significant differences in most of the traits measured. Also, as the NaN₃ concentration increased, germination, flowering, maturity and other yield related traits decreased. However, germination inhibition increased as NaN₃ concentration increased. Hence, NaN₃ was effective in inducing earliness to flowering and maturity, but it hampers seed and tuber yields. TSS 79 accession produced the highest seed yield, when treated with 0.04% concentration of NaN₃, but other genotypes produced very poor seed yields when treated, hence TSS 79 accession is regarded as a good genotype for recommendation for further improvement in mutation studies for increased seed yield. Therefore, NaN₃ is capable of inducing variability in AYB genotypes and can therefore be exploited for the genetic improvement of AYB in breeding programmes.

Keywords: African yam bean, sodium azide, flowering, maturity, seed yield, tuber yield

African yam bean (*Sphenostylis stenocarpa*) (AYB) is one of the under-utilized legumes grown in Nigeria, Central African Republic, Gabon, Zaire and Ethiopia. It produces two consumable parts: the tuber which grows as the root sink and the actual yam beans (seeds), which develop in pods above the ground (Olasoji et al. 2011). The seeds and tubers are of economic importance providing food for humans and livestock. AYB is a good source of protein, containing approximately 29% and 19% crude protein in its grains and tubers, respectively (Adewale and Dumet 2011). The amino acid (lysine and methionine) content in AYB grain is higher than that in pigeon pea, cowpea and bambara groundnut (Uguru and Madukaife 2001). Global food security is being threatened with increasing dependence on a few major staple crops such as cowpea, maize, rice etc. (Ikhajagbe and Mensah 2012). AYB is a potential crop to mitigate the

current alarming reduction in crop diversity and variability, but the present climate change associated with erratic rainfall patterns and high incidence of pests and diseases in the ecosystem, is contributing to low grain yield of AYB in Nigeria. Although, AYB seems to be capable of reducing malnutrition, most of the known accessions are late maturing, as it attains physiological maturity between 6 – 7 months after planting, depending on cultivars (Akinyosoye et al. 2017). It therefore becomes important to breed for early maturing and high yielding varieties that will be adaptable to the various agro-ecologies of Nigeria.

Mutation breeding has been recognized as a valuable supplement to conventional breeding and it has contributed significantly to crop improvement. Presently, 3220 mutant cultivars have been developed in over 220 crop species through mutation and released to

Impact of sodium azide on maturity, seed and tuber yields in M₁ African yam bean; *Solomon Tayo Akinyosoye et al.* farmers all over the world (Bado et al. 2015). Chemical mutagenesis (the non-GMO approach) is a simple approach to create mutation in plants for the improvement of agronomic traits (Roychowdhury and Tah 2011). Sodium azide (NaN₃) has been regarded as one of the most powerful mutagens in crop plants and the mutant plants produced by the treatment of sodium azide are capable of surviving under various adverse conditions and have improved yields, increased stress tolerance, longer shelf-life and reduced agronomic input, in comparison to their normal parents (Khan et al. 2009).

Presently, there is no known registered and released variety of AYB in Nigeria. Some accessions are only found in the hands of old farmers (Saka et al. 2004; Adewale et al. 2012; Ojuederie et al. 2014), thus making it vulnerable to extinction. Also, information regarding research activities on the use of mutation breeding for the genetic improvement of legumes, especially AYB is scarce in Nigeria. This study therefore, evaluated the effects of the mutagen (sodium azide) on maturity, tuber and seed yield related traits of African yam bean accessions, with the aim of creating genetic variation among the AYB genotypes

Materials and methods

Selection of AYB accessions for the experiment

Six promising AYB accessions selected for their earliness to maturity and high yield traits/ characteristics from previous evaluations (unpublished) across locations in southwest Nigeria were selected for the study. The list of the accessions and source of collection are presented in Table 1.

Table 1: List of the accessions of African yam bean and source of collection

Accession name	Source
AYB 94	IAR&T
AYB 61	IAR&T
TSS 79	IITA
TSS 62	IITA
TSS 3	IITA
NGB01349	NACGRAB

IAR&T: Institute of Agricultural Research and Training, Ibadan, Nigeria; IITA: International Institute for Tropical Agriculture, Ibadan, Nigeria; NACGRAB: National Centre for Genetic Resources and Biotechnology, Ibadan, Nigeria

Seed treatment

For each AYB genotype, 500 dry seeds (12% moisture content) were divided into five parts to obtain 100 seeds per treatment. The seeds were soaked in distilled water at room temperature ($25 \pm 2^{\circ}\text{C}$) for 6 hours to soften the seed, thereafter excess water was drained. Seeds were then submerged in solutions containing four different concentrations of sodium azide (NaN₃), 0.02%, 0.04%, 0.06% and 0.08% for 9 hours. The untreated seeds (0%) were also submerged in only distilled water for 9 hours to serve as a control (Sheikh et al. 2012; Raina et al. 2020). A phosphate buffer was used to maintain the sodium azide pH at 3.7. Thereafter, seeds were thoroughly washed in running tap water to remove the residual mutagen from the seed surfaces and then spread on and covered with Whatman 1TM filter paper, so as to surface dry the seeds (Akinyosoye 2020).

Field evaluation

M₁ generation was evaluated during the 2020 cropping season at the experimental field of Institute of Agricultural Research and Training, Obafemi Awolowo University, Moor Plantation, Ibadan, Nigeria.

The experiment was laid out in a randomized complete block design with three replications. The plot size was 5 m by 5 m and the crops were planted at the spacing of 1 m by 1 m between and within the rows, respectively. Two seeds were planted per stand with the aim of having 25 stands per plot and 50 plants per plot. Weeds were manually managed. Pest infestations were first noticed on the plants at the reproductive stage and were controlled by using cypermethrin with active ingredient: (200 g/l pyrethroid) E.C (NPIC, 1988) mixed in a knapsack sprayer with fungicide powder benlate, with active ingredient 50% methyl 1-(butylcarbamoyl-2-benzimidazolecarbamate), manufactured by DuPont company, Wilmington, Delaware, U.S.A. Data were collected on germination characters are as follows:

Percentage (%) germination of each plot was calculated:

$$\frac{\text{Number of germinated seed}}{\text{Total number of seeds planted}} \times 100$$

Inhibition germination percentage was determined:

$$\frac{\% \text{Germination of untreated} - \% \text{Germination of treated}}{\% \text{Germination of untreated}} \times 100$$

Data were also collected on days to first flowering, days to 50 % flowering, and days to 70% physiological maturity to determine the time to maturity. Yield related traits (number of seeds per pod, number of pods per peduncle, number of pods per plant, number of pods per peduncle, pod weight per plant, pod length, seed yield per plant and tuber yield per plant) were measured according to the guide on AYB descriptors (Adewale and Dumet 2011). Data obtained were subjected to analyses of variance (ANOVA). Differences between the treatments were separated using Duncan Multiple Range Test at 5 % or 1 % levels of significance. Pearson

correlations between pair of seed and tuber yields with seed germination traits and other agronomic traits were estimated using Statistical Tool for Agricultural Research (version 2.0.1, 2013 – 2020).

Results and discussion

Analysis of variance for seed germination characteristics, flowering, maturity and yield related characteristics in AYB as affected by genotype, varied concentrations of sodium azide and their interactions

Table 2 shows the seedling germination traits, flowering, maturity, seed and tuber yields of African yam bean (AYB) genotypes were significantly different ($P \leq 0.01$). Also, varied concentrations of NaN_3 had significant effect on all the agronomic and yield related traits measured in this study, excepted days to 50% flowering. Similarly, interactive effect between genotypes and varied concentrations of NaN_3 were significant for most of the traits measured, with the exception of peduncle length, number of peduncles, pods per plant, pod length and 70% physiological maturity. This indicates that performance of the AYB for most of the traits measured in this study may be attributed to their inherent genetic make-up, varied NaN_3 concentrations and their interactions. This is in agreement with the report of others on different crops (Wannajindaporn et al. 2014; Akinyosoye 2020).

Table 2 also shows that a high degree of genotypic variations existed in most of yield components among the AYB genotypes evaluated in this study as reflected by the coefficients of variation (CV), where pod length had the highest CV (113.1%), while the lowest CV was obtained in days to 50% flowering (3.8%). Thus, the high CVs (> 50%) obtained in seed yield per plant and in most of

Impact of sodium azide on maturity, seed and tuber yields in M₁ African yam bean; *Solomon Tayo Akinyosoye et al.* yield components, would facilitate effective selection for grain yield improvement in AYB mutation breeding programmes, whereas low CVs (<50%) obtained in all of seed germination characteristics, flowering and maturity in this study, pointed out that aforementioned traits could be further improved through introduction, selection, hybridization and recombination.

Table 2: Combined mean squares for seed germination parameters, flowering, maturity and yield related characteristics in African yam bean as affected by genotype, varied concentration of sodium azide and their interactions

	Df	GP%	PGI%	DFP	D50F	MAT	SYPP (g)	TYPP (g)	POPEP	PEPP	POPP	POL (cm)	SPPO	POWPP (g)
Genotype (G)	5	4217.11**	3859.71**	9284.17**	10389.50**	440.02**	321.87**	4790.77**	8.55**	55.36**	238.88**	424.21**	145.90**	1550.17**
Concentration (C)	4	13084.44**	21818.38**	9598.73**	10566.04	558.40**	755.18**	4679.38**	3.81**	47.77**	173.32**	294.17*	127.09**	2021.35**
G x C	20	267.11*	549.31**	2991.26**	3275.50**	8.32	170.37**	1722.91**	1.07**	8.88	33.61	96.63	35.58*	493.90*
Error	58	152.14	188.28	14.29**	12.73**	13.00	47.13	270.95	0.69	7.07	22.94	77.95	20.51	229.47
Mean		33.20	59.82	106.54	111.73	184.00	6.49	16.75	1.61	5.22	9.59	7.81	5.19	14.09
SE(0.05)		1.30	1.43	0.39	0.37	0.38	0.712	1.76	0.087	0.29	0.52	0.94	0.48	1.59
CV%		38.41	22.38	4.23	3.81	4.50	105.86	98.27	52.01	50.97	49.96	113.11	87.27	107.52

*, ** Significant at (P≤0.05) and (P≤ 0.01) respectively; GP%: percentage germination; PGI%: percentage germination inhibition; DFP: days to first flowering; D50F: days to 50% flowering; MAT: days to 70% physiological maturity; SYPP: seed yield per plant; TYPP: tuber yield per plant; POPEP: number of pods per peduncle; PEPP: number of peduncles per plant; POPP: number of pods per plant; POL: pods length; SPPO: number of seeds per pod; POWPP: pod weight/plant; CV: coefficient of variation

Effects of genotype and varied concentration of sodium azide on seed germination traits, flowering and maturity

Table 3 shows there were genotypic differences in the mean performance of the AYB genotypes for seed germination traits, flowering and maturity. TSS 79 had highest germination percentage (52.67%), with AYB 94 (45.33%) and AYB 61 (43.33%) also having significantly higher germination percentages than the other genotypes. Two genotypes (NGB01349 and TSS 3) flowered and matured earlier than the other genotypes across treatments, both these genotypes attained first flowering within 104–105 days after seeding (DAS), 50% flowering within 108–109 DAS and both reached 70% physiological maturity in 177 DAS. Therefore,

genotypic differences for germination traits, flowering and maturity period of the AYB genotypes could be linked to the effects of varied concentration of sodium azide imposed on them.

Effect of varied concentration of NaN₃ showed that as the concentration of NaN₃ increased, germination percentage decreases from 0% to 0.08% concentration, whereas percentage germination of inhibition increased as the concentration of NaN₃ increases (Table 3). The results obtained in this study had been reported in various crops, including maize (Akinyosoye 2020); cowpea (Gnanamurthy and Dhanavel 2014); soybean (Dhanavel et al. 2008); tomato (Adebola 2013); and wheat (Srivastava et al. 2010). The reduction in seed germination and increase inhibition in seed germination in M₁ African yam bean

generation due to sodium azide in this study may be attributed to changes in physiological, biochemical and biological processes as the concentration of the sodium azide increases (Akinyosoye 2020). Salim et al. (2009); Srivastava et al. (2011) reported that NaN₃ effect hampered adenosine triphosphate (ATP) biosynthesis resulting in decreased ATP molecules. Thus, all living cells require energy in the form of ATP molecules to perform biological processes, but at low energy; could lead to decrease in ATP molecules availability and translates to reduction in germination rate or growth and development.

Table 3 shows that, varied concentrations of NaN₃ induced early flowering and

physiological maturity as the AYB genotypes treated with NaN₃, flowered and attained 70% physiological maturity earlier than the control in this study. Induction of earliness by NaN₃ in the African yam bean evaluated in this study, could be because induced mutation is non-specific as it is a random event, might possibly affect the gene(s) responsible for flowering and maturity as well as yield by switching from yield production to early flowering and physiological maturity; or vice-versa or favour both at the same time. Induction of early flowering in some crops treated with mutagenic agents has been reported (Mensah et al. 2007; Bolbhat and Dhumal 2012; Animasaun et al. 2014).

Table 3: Means of seed germination traits, flowering and maturity as affected by genotype and varied concentration of sodium azide

Accessions	Germination percentage (%)						Percentage germination Inhibition (%)					
	0.0%	0.02%	0.04%	0.06%	0.08%	Mean	0.0%	0.02%	0.04%	0.06%	0.08%	Mean
AYB 61	90.00	26.67	33.33	40.00	26.67	43.33a	0.00	70.37	62.96	55.56	70.37	51.85b
AYB 94	86.67	53.33	53.33	13.33	20.00	45.33a	0.00	39.47	35.87	85.19	76.40	47.39b
NGB01349	66.67	6.67	6.67	6.00	4.00	18.00b	0.00	90.48	90.48	92.00	94.00	73.39a
TSS 3	76.67	13.33	7.00	6.67	3.00	21.33b	0.00	82.14	91.00	90.48	97.00	72.12a
TSS 62	66.67	6.67	6.67	6.00	6.67	18.54b	0.00	90.48	90.00	91.50	90.48	72.49a
TSS 79	90.00	60.00	40.00	40.00	33.33	52.67a	0.00	33.52	56.11	55.19	63.52	41.67b
Mean	79.45a	27.78b	24.50b	18.67b	15.61b		0.00c	67.74b	71.07ab	78.32a	81.96a	
	Days to first flowering						Days to 50% flowering					
AYB 61	113.00	103.67	107.67	105.67	103.33	106.67a	117.00	106.33	111.67	112.00	109.00	111.20a
AYB 94	117.67	109.67	110.33	106.67	105.00	109.87a	121.67	117.33	116.00	114.00	115.00	116.80a
NGB01349	108.33	106.00	106.00	99.00	100.00	103.87b	112.33	110.00	109.00	105.00	102.00	107.67b
TSS 3	109.00	107.00	106.00	104.00	98.67	104.93b	111.00	109.00	109.00	111.00	104.33	108.87b
TSS 62	110.00	110.00	105.00	105.00	110.00	108.00a	117.67	117.00	107.00	109.00	117.00	113.53a
TSS 79	109.33	106.33	105.67	105.00	103.33	105.93a	121.33	116.67	108.67	107.00	108.00	112.33a
Mean	111.22a	107.11b	106.78b	104.22b	103.39b		116.83a	112.72a	110.22a	109.67a	109.22a	
	Days to 70% physiological maturity											
AYB 61	201.00	186.00	186.00	186.00	186.00	189.00a						
AYB 94	201.00	186.00	186.00	186.00	186.00	189.00a						
NGB01349	186.00	177.00	177.00	174.00	170.00	176.80b						
TSS 3	186.00	177.00	177.00	175.00	171.00	177.20b						
TSS 62	190.00	180.00	185.00	180.00	180.00	183.00a						
TSS 79	201.00	186.00	186.00	186.00	186.00	189.00a						
Mean	194.17a	182.00b	182.83b	181.17b	179.83b							

Means with the same letter(s) in the same column or row are not significantly different from each other at P > 0.05

Impact of sodium azide on maturity, seed and tuber yields in M₁ African yam bean; *Solomon Tayo Akinyosoye et al.*
Effects of genotype and varied concentration of sodium azide on yield related traits

Genotypic differences also existed in the mean performance of the AYB genotypes for seed and tuber yields as well as yield related traits in this study. Table 4 shows that the genotype TSS 79 recorded the highest seed yield (13.70 g) per plant across treatments. Also, TSS 79 recorded the highest seed yield of 19.67 g per plant when treated with 0.04% concentration of NaN₃, when compared with the control (14.83 g). Two genotypes (AYB 61 and AYB 94) had highest tuber yields with mean values of 38.07 g and 39.00 g per plant respectively, across treatments. The same genotypes (AYB 61 and AYB 94) recorded highest tuber yield of 132.67 g and 94.67 g per plant respectively, in the control treatment, but AYB 61 gave no yields when treated with 0.08% concentration of NaN₃. For the yield related traits, TSS 79 performed better than other genotypes across treatments in this study. The observed genotypic differences had been reported by some researchers who obtained higher yield at a lower dose of sodium azide (Khan and Goyal 2009). Hence, sodium azide has been reported to have caused positive and negative shifts in yield related traits in wheat genotypes (Srivastava et al. 2010).

Effect of varied concentration of NaN₃ on seed and tuber yields per plant showed that the control had the highest mean values of 17.70 g and 44.47 g for seed and tuber yields, respectively, but these decreased to 6.11 g and 12.39 g for seed and tuber yields per plant, respectively at 0.08% concentration. Consequently, other yield contributing traits

followed the same trends as observed in both seed and tuber yields. The results obtained are in agreement with the work of Kumar et al. (2009) who reported that chemical mutagens induced physiological damages (injury), gene mutations and chromosomal mutations in the organisms in M₁ generation in terms of lethality, fertility reduction or sterility (reduction in pods and seed yield). Hence, Adamu et al. (2004) suggested that the trend in some characteristics observed in their studies were concentration dependent.

During the experiment it was observed that leaves and tuber size of AYB plants in the field revealed that plant stands of TSS 79 in control treatment had multiple infections, characterized by leaves lesion, yellowish and wrinkled leaves, especially at the reproductive stage. To combat this the plants were sprayed with insecticide and fungicide, this could be responsible for the low seed yield obtained in the control treatment of TSS 79 when compared with the 0.04% concentration (Table 4; Figure 1A). However, it was observed that varied concentration of NaN₃ affected the tuber formation and development, and all of the genotypes in control treatments produced tubers with big sizes, whereas some of the genotypes treated with NaN₃ had tubers with small sizes and some did not produce tubers (Table 4; Figure 1C and 1D). This could be due to inherent genetic potentials of AYB genotype to varied concentrations of NaN₃ and effectiveness of NaN₃ against field pests. This observation corroborated by the findings of Rodriguez-Kabana and Akridge (2003) who reported that application of NaN₃ on green pepper was capable of controlling weeds and soil-borne pests.

Table 4: Means of yield related traits as affected by genotype and varied concentration of sodium azide

	Seed yield per plant (g)						Tuber yield per plant (g)					
	0%	0.02%	0.04%	0.06%	0.08%	Mean	0%	0.02%	0.04%	0.06%	0.08%	Mean
AYB 61	0.67	0.33	4.67	1.67	2.67	2.00d	132.67	52.00	5.67	0.00	0.00	38.07a
AYB 94	40.39	1.00	0.67	0.00	0.00	8.41bc	94.67	27.33	20.00	20.00	33.00	39.00a
NGB01349	10.33	1.00	0.00	0.00	0.00	2.27d	4.67	3.00	0.00	0.00	20.00	5.53bc
TSS 3	25.67	0.00	0.00	2.00	16.67	8.87ab	3.00	0.00	0.00	0.00	0.00	0.60c
TSS 62	14.33	0.00	0.00	0.00	4.00	3.67cd	3.00	0.00	0.00	0.00	0.00	0.60c
TSS 79	14.83	12.33	19.67	8.33	13.33	13.70a	28.83	12.33	4.00	17.00	21.33	16.70b
Mean	17.70a	2.44b	4.17b	2.00b	6.11b		44.47a	15.78b	4.94b	6.17b	12.39b	
	Pod length (cm)						Number of seeds per pod					
AYB 61	7.67	6.33	13.50	6.67	15.33	9.90b	4.67	4.00	8.83	4.00	7.33	5.77bc
AYB 94	11.31	13.67	5.22	0.00	0.00	6.04bc	7.06	5.00	2.83	0.00	0.00	2.98c
NGB01349	14.67	5.33	0.00	0.00	0.00	4.00c	10.67	3.33	0.00	7.06	5.00	5.21ab
TSS 3	24.67	0.00	0.00	4.33	16.20	9.04b	2.83	0.00	0.00	5.33	12.50	4.13bc
TSS 62	6.67	0.00	0.00	0.00	0.00	1.33c	9.00	0.00	0.00	0.00	3.33	2.47c
TSS 79	20.75	15.90	7.33	17.35	21.28	16.52a	11.50	10.50	10.33	10.50	9.67	10.50a
Mean	14.29a	6.87b	4.34b	4.73b	8.80ab		9.70a	3.81b	3.67b	3.31b	5.47b	
	Number of pods per peduncle						Number of pods per plant					
AYB 61	2.60	1.33	2.17	2.67	2.67	2.29a	14.33	9.00	12.83	14.14	12.00	12.46a
AYB 94	2.43	2.89	1.67	1.33	2.33	2.13a	13.40	12.56	12.44	10.00	13.00	12.28a
NGB01349	2.00	1.33	1.33	0.00	1.33	1.20b	14.87	7.00	9.33	0.00	4.00	7.04b
TSS 3	2.17	0.67	0.00	0.67	2.17	1.13bc	15.22	9.67	4.33	0.00	13.25	8.49b
TSS 62	2.00	0.67	0.00	0.00	0.00	0.53c	13.00	4.00	0.00	0.00	0.00	3.40c
TSS 79	2.62	2.17	2.00	2.70	2.33	2.36a	17.15	13.89	14.00	12.87	11.33	13.85a
Mean	2.30a	1.51bc	1.19c	1.23bc	1.81ab		14.66a	9.35b	8.82b	6.17b	8.93b	
	Number of peduncles per plant						Pod weight per plant (g)					
AYB 61	7.93	5.33	7.17	7.53	6.83	6.96a	1.33	0.67	20.83	5.33	7.00	7.03b
AYB 94	7.57	6.56	6.33	5.17	6.83	6.49ab	48.67	8.00	4.17	0.00	0.00	12.17b
NGB01349	7.83	3.33	5.33	0.00	2.00	3.70c	29.67	1.67	0.00	0.00	0.00	6.27b
TSS 3	8.33	5.00	2.33	0.00	7.25	4.58bc	49.33	0.00	0.00	3.00	32.50	16.97b
TSS 62	6.50	2.67	0.00	0.00	3.20	2.47c	37.00	0.00	0.00	0.00	7.33	8.87b
TSS 79	8.25	7.61	7.00	6.67	6.00	7.11a	29.50	30.17	27.67	46.67	32.17	33.23a
Mean	7.74a	5.08bc	4.69bc	3.23c	5.35b		32.58a	6.75b	8.78b	9.17b	13.17b	

Means with the same letter(s) in the same column or row are not significantly different from each other at $P > 0.05$



Figure 1A: Diseased leaves of TSS 79 at reproductive stage (control or untreated with NaN₃), before spraying with insecticide and fungicide in the field



Figure 1B: Smooth and disease-free leaves of TSS 79 at reproductive stage (0.04% concentration of NaN₃)



Tubers of AYB 94 with highest tuber weights in the control treatment



Tubers of AYB 94 at harvest (0.08% concentration of NaN₃)

Figure 1: Visual assessment leaves and tuber size of African yam bean in control (untreated) and treatment with Sodium azide (NaN₃)

Correlation between pairs of tuber and seed yields with other yield contributing traits

Pearson correlation between tuber yield per plant and seed yield revealed that a non-significant association existed between them ($r = 0.15$) (Table 5). This result corroborates the report of Akinyosoye et al. (2017) who

reported non-significant and positive correlation between seed and tuber yields in AYB. This phenomenon could be due to competition between the storage sinks (seeds and tubers) to import photo-assimilates, which is a function of environmental factors and strength of the competing sinks (Farrar 1996; Geigner et al. 1996).

Tuber yield was positively and significantly correlated with percentage

Impact of sodium azide on maturity, seed and tuber yields in M₁ African yam bean; *Solomon Tayo Akinyosoye et al.*
 germination ($r = 0.49^{**}$), first flowering ($r = 0.28^{**}$), 50% flowering ($r = 0.28^{**}$), 70% physiological maturity ($r = 0.66^{**}$), number of pods per peduncle ($r = 0.28^{**}$), number of pods per plant ($r = 0.24^*$), and number of peduncle per plant ($r = 0.25^*$), but negatively and significantly correlated with percentage germination inhibition ($r = -0.42^{**}$). This suggests that high germination percentage, late flowering and maturity, increase in number of pods per peduncle, pods per plant and peduncle per plant contributed to tuber yield. But percentage germination inhibition hampers tuber formation.

Positive and significant relationships were obtained between seed yield per plant and first flowering ($r = 0.24^*$), 50% flowering ($r =$

0.23^*), 70% physiological maturity ($r = 0.40^{**}$), pod length ($r = 0.46^{**}$), number of seeds per pod ($r = 0.56^{**}$), number of pods per peduncle ($r = 0.31^{**}$), number of pods per plant ($r = 0.31^{**}$), number of peduncle per plant ($r = 0.30^{**}$) and pod weight per plant ($r = 0.86^{**}$), but negatively and significantly associated with percentage germination inhibition ($r = -0.49^{**}$). Thus, all positive and significant among yield related traits with seed yield, could be used to improve seed yield. Bello and Olawuyi (2015) reported that significant positive correlations between yield and other yield related traits can improve yield, which are quite desirable in plant breeding, because it facilitates the selection process and gains from selection.

Table 5: Pearson correlation coefficients between pairs of tuber and seed yields with seed germination parameters, earliness and other yield related traits

	TYPP	PSG%	PGI%	DFF	D50F	MAT	POL	SPPO	POPEP	POPP	PEPP	POWPP	SYPP
TYPP	1	0.49**	-0.42**	0.28**	0.28**	0.66**	0.05	-0.02	0.28**	0.24*	0.25*	0.03	0.15
PSG%		1	-0.98**	0.46**	0.47**	0.83**	0.47**	0.49**	0.58**	0.63**	0.60**	0.49**	0.51**
PGI%			1	-0.46**	-0.47**	-0.79**	-0.43**	-0.48**	-0.54**	-0.63**	-0.59**	-0.49**	-0.49**
DFF				1	0.99**	0.39**	0.27**	0.24*	0.57**	0.64**	0.59**	0.26*	0.24*
D50F					1	0.40**	0.28**	0.25*	0.57**	0.65**	0.59**	0.26*	0.23*
MAT						1	0.31**	0.29**	0.50**	0.49**	0.47**	0.34**	0.40**
PODL							1	0.80**	0.46**	0.40**	0.39**	0.62**	0.46**
SPPO								1	0.41**	0.42**	0.39**	0.67**	0.56**
POPEP									1	0.82**	0.77**	0.39**	0.31**
POPP										1	0.95**	0.35**	0.31**
PEPP											1	0.33**	0.30**
POWPP												1	0.86**
SYPP													1

TYPP: tuber yield per plant; PSG%: percentage seed germination; PGI%: percentage germination inhibition; DFF: days to first flowering; D50F: days to 50% flowering; MAT: POL: pods length; days to 70% physiological maturity; SPPO: number of seeds per pod; POPEP: number of pods per peduncle; POPP: number of pods per plant; PEPP: number of peduncles per plant; POWPP: pod weight/plant; SYPP: seed yield per plant

Conclusion

Sodium azide was effective in inducing earliness to flowering and maturity, but it hampers seed and tuber yields. TSS 79 genotype produced the highest yield, when treated with 0.04% concentration of sodium azide, but other genotypes produced very poor seed yields when treated, hence TSS 79 is regarded as a good genotype to be recommended for further improvement through mutation studies for increased seed yield. Sodium azide is capable of inducing variability in African yam bean genotypes and can therefore be exploited for the genetic improvement of African bean in breeding programs.

References

- Adamu, A.K., S.S. Clung, and S. Abubakar. 2004. "Effects of Ionizing Radiation (Gamma rays) on Tomato (*Lycopersicon esculentum* L.)." *Nigerian Journal of Experimental and Applied Biology* **5(2)**: 185–193.
- Adebola, M.O. 2013. "Mutagenic Effects of Sodium Azide (NaN₃) on Morphological Characteristics of Tomato (*Lycopersicon esculentum*)." *International Journal of Research Publication* **2 (4)**: 1563–2251.
- Adewale, B.D., D.J. Dumet, I. Vroh-Bi, O.B. Kehinde, D.K. Ojo, A.E. Adegbite, and J. Franco. 2012. "Morphological Diversity Analysis of African Yam bean and Prospects for Utilization in Germplasm Conservation and Breeding." *Genetic Resources and Crop Evolution* **59(5)**: 927–936. doi:10.1007/s10722-011-9734-1
- Adewale, B.D., and D.J. Dumet. 2011. "Descriptors for African Yam Bean *Sphenostylis stenocarpa* (Hochst ex. A. Rich.) Harms." Available at: http://old.iita.org/cms/articulatefiles/1488-ayb_descriptors.
- Akinyosoye, S.T. 2020. "Effect of Sodium Azide (NaN₃) on Seed Germination and Callus Induction in Some Maize (*Zea mays* L.) Varieties." *Nigerian Journal of Seed Science* **4**:66–75.
- Akinyosoye, S.T., J.A. Adetumbi, O.D. Amusa, O.A. Agbeleye, F.B. Anjorin, M.O. Olowolafe, and T. Omodele. 2017. "Bivariate Analysis of the Genetic Variability among Some Accessions of African Yam Bean (*Sphenostylis stenocarpa* Hochst Ex A. Rich)." *Acta Agriculturae Slovenica* **109**:493–507.
- Animasaun, D.A., S. Oyediji, M.A. Azeez, and A.O. Onasanya. 2014. Alkylating Efficiency of Sodium Azide on Pod Yield, Nut Size and Nutrition Composition of Samnut 10 and Samnut 20 Varieties of Groundnut (*Arachis hypogea* L.)." *African Journal of Food Agriculture Nutrition and Development* **14**:9497–9510.
- Bado, S., B.P. Forster, S. Nielen, A. Ghanim, P.J.L., Lagoda, B.J. Till, and M. Laimer. 2015. "Plant Mutation Breeding: Current Progress and Future Assessment." *Plant Breeding Reviews* **39**:23–88. <http://dx.doi.org/10.1002/9781119107743.ch02>.
- Bello, B.O., and O.J. Olawuyi. 2015. "Gene Action, Heterosis, Correlation and Regression Estimates in Developing Hybrid Cultivars in Maize." *Tropical Agriculture (Trinidad)* **92 (2)**: 102–117.
- Bolbhat, S.N., and K.N. Dhumal. 2012. "Effect of Mutagens on Quantitative Characters in M₂ and M₃ Generation of Hoersegram (*Macrotyloma uniflorum* (Lam.) Verdc)." *International Journal of Scientific and Research Publication* **2(10)**: Accessed Oct, 2012 www.ijsrp.org
- Dhanavel, D., P. Pavadai, L. Mullainathan, D. Mohana, G. Raju, M. Girija, and C. Thilagavathi. 2008. "Effectiveness and Efficiency of Chemical Mutagens in Cowpea (*Vigna unguiculata* L.) Walp.)." *African Journal of Biotechnology* **7**:4116–4117.
- Farrar, J.F. 1996. "Sink-Integral Parts of a Whole Plant." *Journal of Experimental Botany* **47**:1273–1279. doi:10.1093/jxb/47.

- Impact of sodium azide on maturity, seed and tuber yields in M₁ African yam bean; *Solomon Tayo Akinyosoye et al.*
Special_Issue.1273
- Geigner, D., K.E. Koch, and W.J. Shieh. 1996. "Effect of Environmental Factors on Whole Plants Assimilates Partitioning Associated Gene Expression." *Journal of Experimental Botany* **47**:1229–1238. doi:10.1093/jxb/47. Special Issue.1229.
- Gnanamurthy, S., and D. Dhanavel. 2014. "Effect of EMS on Induced Morphological Mutants and Chromosomal Variation in Cowpea (*Vigna unguiculata* (L.) Walp)." *International Letters of Natural Sciences* **17**:33–43.
- Ikhajagbe, B., and J.K. Mensah. 2012. "Genetic Assessment of Three Colour Variants of African yam Beans (*Sphenostylis stenocarpa*) Commonly Grown In Mid-Eastern Region of Nigeria." *International Journal of Modern Botany* **2** (2): 13–18. doi:10.5923/j.ijmb.20120202.01
- Khan, S., F. Al-Qurainy, and F. Anwar. 2009. "Sodium Azide: A Chemical Mutagen for Enhancement of Agronomic Traits of Crop Plants." *Environ Int J Sci Tech* **4**:1–21.
- Khan, S., and S. Goyal. 2009. "Improvement of Mungbean Varieties through Induced Mutations." *African Journal of Plant Science* **3** (8): 74–180.
- Kumar, V.A., R.U. Kumari, R. Amutha, T.S. Kumar, S.J. Hepziba, and C.R.A. Kumar. 2009. "Effect of Chemical Mutagen on Expression of Characters in Arid Legume Pulse-Cowpea (*Vigna unguiculata* L. Walp.)." *Research Journal of Agriculture and Biological Sciences* **5** (6): 1115 – 1120.
- Mensah, J.K., B.O. Obadoni, P.A. Akomeah, B. Ikhajagbe, and J. Ajibulu. 2007. "The Effects of Sodium Azide and Colchicine Treatments on Morphological and Yield Traits of Sesame Seed (*Sesame indicum* L.)." *African Journal of Biotechnology* **6**(5): 534–538.
- NPIC (National Pesticide Information Centre). 1988. Factsheets. Corvallis: Oregon State University and the United States Environmental Protection Agency.
- Ojuederie, O.B., M.O. Balogun, I. Fawole, O.I. David, and M.O. Olowolafe. 2014. "Assessment of the Genetic Diversity of African Yam Bean (*Sphenostylis stenocarpa* Hochst ex. A Rich. Harms) Accessions Using Amplified Fragment Length Polymorphism (AFLP) Markers." *African Journal of Biotechnology* **13** (18): 1850–1858.
- Olasoji, J., S. Akande, and O. Owolade. 2011. "Genetic Variability in Seed Quality of African Yam Bean (*Sphenostylis stenocarpa* Hoscht. Ex A. RichHarms)." *African Journal of Agricultural Research* **6** (27): 5848–5853.
- Raina, A., R.A. Laskar, Y.R. Tantray, S. Khursheed, M.R. Wani, and S. Khan. 2020. "Characterization of Induced High Yielding Cowpea Mutant Lines using Physiological, Biochemical and Molecular Markers." *Scientific Report* **10**: 3687. doi.org/10.1038/s41598-020-60601-6
- Rodriguez-Kabana, R., and J. R. Akridge. 2003. "Sodium Azide [SEP 100] for Control of Nematodes and Weed Problems in Green Pepper Production." *Proceedings Annual International Research Conference on Methyl Bromide Alternatives and Emissions Reductions, San Diego, California, Nov. 3–6, 2003*. Pages 46–1 to 46–8.
- Roychowdhury, R., and J. Tah. 2011. "Assessment of Chemical Mutagenic Effects in Mutation Breeding Programme for M₁ Generation of Carnation (*Dianthus caryophyllus*)." *Research in Plant Biology* **1** (4): 23–32.
- Saka, J.O., S.R. Ajibade, O.N. Adeniyana, R.B. Olowoyo, and B.A. Ogunbodede. 2004. "Survey of Underutilized Grain Legume Production Systems in South-West Agricultural Zone of Nigeria." *Journal of Agriculture and Food Production* **6**:93–108.
- Salim, K., A. Fahad, and A. Firoz. 2009. "Sodium Azide: A Chemical Mutagen for Enhancement of Agronomic Traits of Crop

- Impact of sodium azide on maturity, seed and tuber yields in M₁ African yam bean; *Solomon Tayo Akinyosoye et al.* Plants.” *Environment, We. International Journal of Science and Technology* **4**:1–21.
- Sheikh, S.A., M.R. Wani, M.A. Lone, M.A. Tak, and N.A. Malla. 2012. “Sodium Azide Induced Biological Damage and Variability for Quantitative Traits and Protein Content in Wheat (*Triticum aestivum* L.)” *Journal of Plant Genomics* **2** (1): 34–38.
- Srivastava P., S. Marker, P. Pandey, and D.K. Tiwari, 2010. Mutagenic Effects of Sodium Azide on the Growth Yield Characteristics in Wheat (*Triticum aestivum* L. em. Thell.). *Asian Journal of Plant Science* **10**:190–201.
- Statistical Tool for Agricultural Research Version: 2.0.1, 2013 – 2020. Rice Research Institute (IRRI). Available:<http://bbi.irri.org>.
- Uguru, M.I., and S.O. Madukaife. 2001. “Studies on the Variability in Agronomic and Nutritive Characteristics of African Yam Bean (*Sphenostylisstenocarpa*) (Hochst ex. A. Rich. Harms),” *Plant Prod. Res. J.* **6**:10–19.
- Wannajindaporn, A., O. Poolsawat, W. Chaowiset, and P.A. Tantasawat. 2014. “Evaluation of Genetic Variability in In Vitro Sodium Azide-Induced Dendrobium ‘Earsakul’ Mutants.” *Genet. Mol. Res.* **13**:5333–5342.